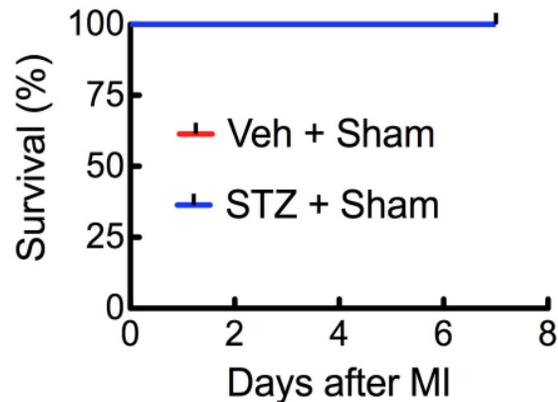
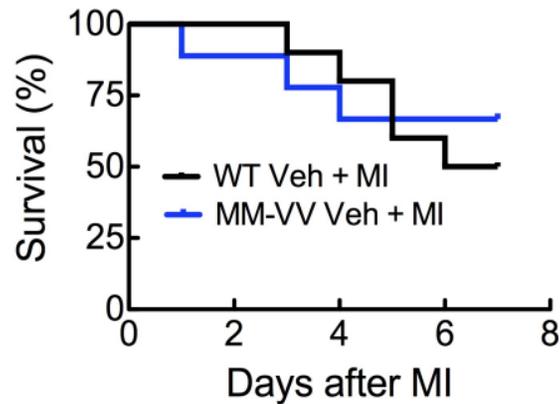


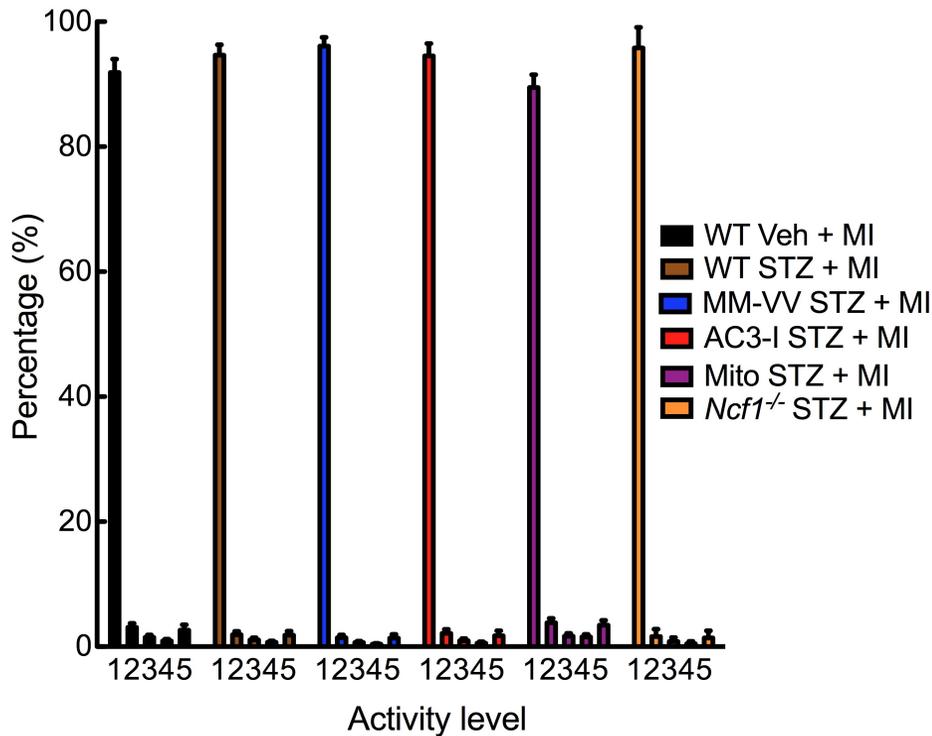
Supplemental Figure 1

STZ causes hyperglycemia. Blood glucose levels after STZ treatment in WT, MM-VV, AC3-I, *Ncf1*^{-/-} and Mito-treated WT mice as well as after Insulin replacement (overall $P < 0.0001$, *** $P < 0.001$, $n = 14-38$ /group). Veh, vehicle; WT, wild type; Mito, MitoTEMPO

A**B**

Supplemental Figure 2

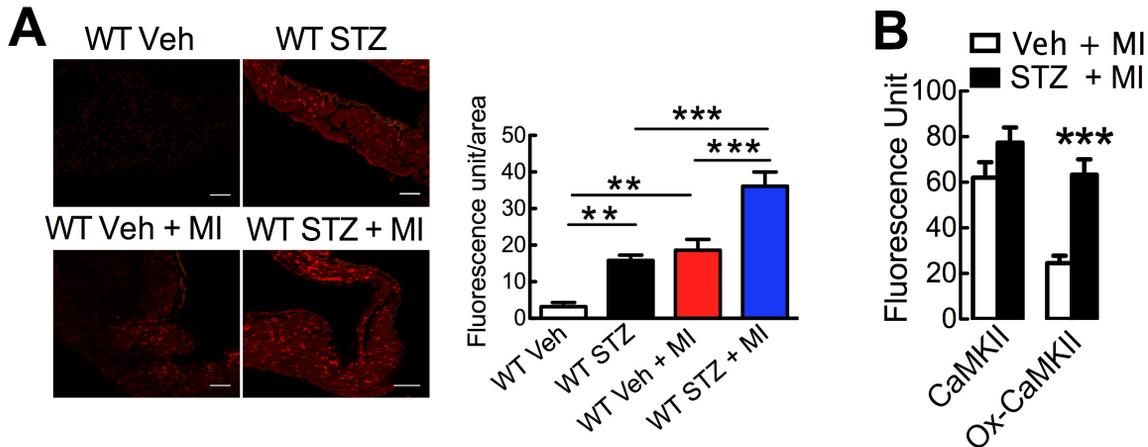
Similar survival after sham surgery in diabetic or non-diabetic mice. (A) Survival in WT STZ- and Veh-treated mice after sham surgery, $P = 1.0$ (the red and blue lines are superimposed), $n = 5/\text{group}$. **(B)** Similar survival after MI in non-diabetic WT and MM-VV mice, $P = 0.7$, $n = 9-10/\text{group}$. WT, wild type; Veh, vehicle.



Supplemental Figure 3

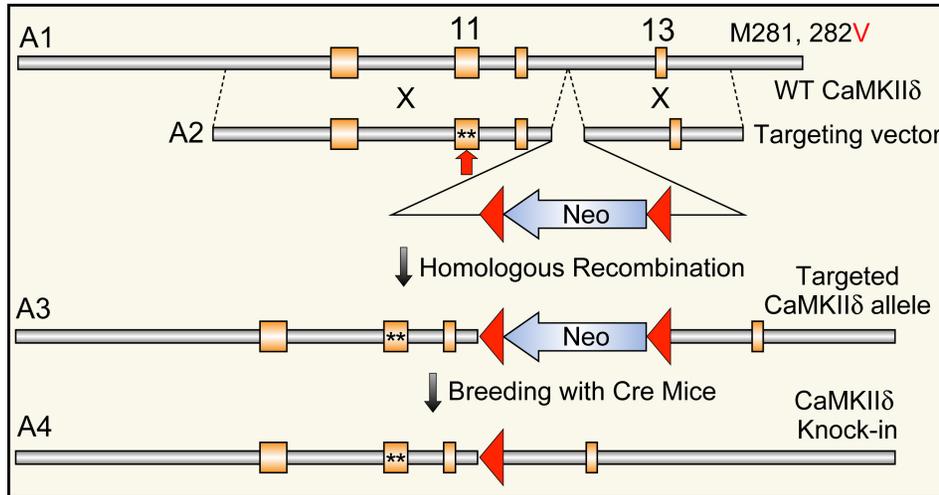
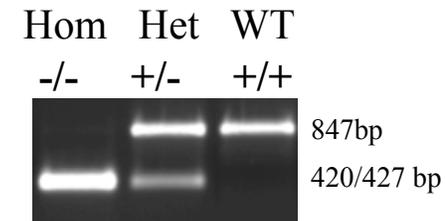
Similar activity levels in diabetic and non-diabetic mice after MI.

Spontaneous activity levels (1-5) between WT, MM-VV, AC3-I, *Ncf1*^{-/-} or WT + Mito mice after STZ + MI. $P = 0.2$ for activity 0 (1), $P = 0.11$ for activity 1-10 (2), $P = 0.4$ for activity 11-15 (3), $P = 0.07$ for activity 16-20 (4), $P = 0.4$ for activity >20 (5), $n = 4-10$ /group. Veh, vehicle; WT, wild type; Mito, MitoTEMPO



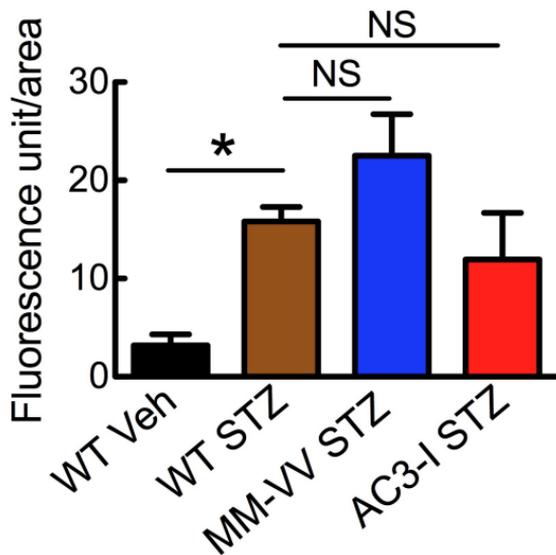
Supplemental Figure 4

Increased ROS and ox-CaMKII in SAN in STZ-treated mice after MI. (A) Representative immunofluorescence images of DHE staining in SAN tissues from WT Veh and WT STZ mice with or without MI. Scale bars: 50 μ m. Overall $P < 0.0001$ by one way ANOVA, $***P < 0.001$, $**P < 0.01$ by Neuman-Keumal's test as post hoc study, $n = 4-6$ /group. (B) CaMKII ($P = 0.14$) and ox-CaMKII ($***P = 0.0004$) immunostaining in SAN tissues from WT Veh and WT STZ with MI ($n = 6$ /group). Veh, vehicle; WT, wild type;

A**B**

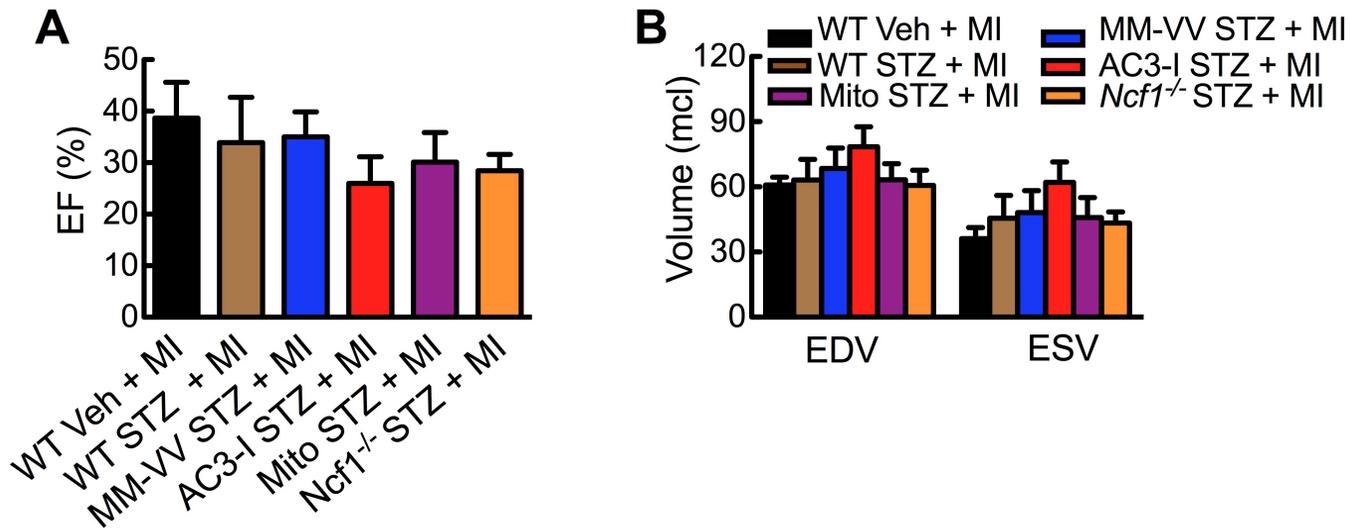
Supplemental Figure 5

Schematic diagram for gene targeting approach for developing MM-VV mice. (A) A partial map of the CaMKII δ gene (A1), the targeting construct (A2), the resulting targeted allele (A3), and the targeted allele after Cre recombination (A4) are shown. In the targeting construct, a 6kb fragment containing exons 9-12 (mutations in Exon 11) and a 4.5-kb fragment containing exon 13 were used as long and short homologous arms, respectively, flanking a LoxP Neo selection marker. (B) The knock-in mouse (F3) genotype. The gel showed the HincII-digested PCR products of homozygote (Hom, -/-), heterozygote (Het, +/-) and wild-type (WT, +/+) mice.



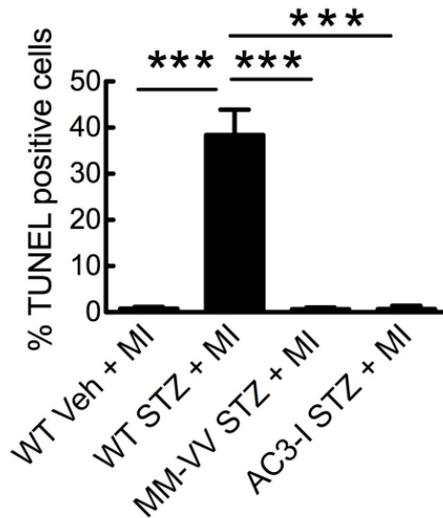
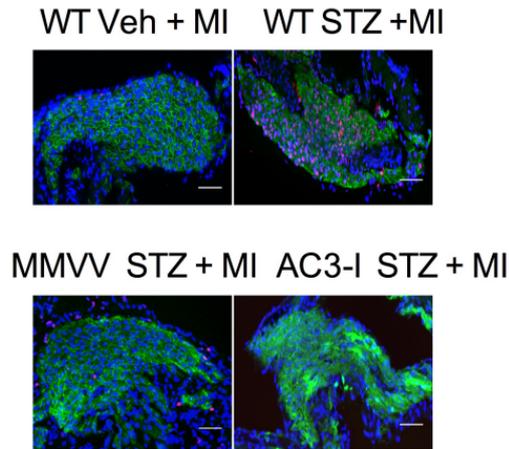
Supplemental Figure 6

ROS in SAN from MM-VV, AC3-I and WT diabetic mice. DHE staining in SAN tissues from Veh- and STZ-treated WT mice, STZ-treated MM-VV and AC3-I mice. Overall $P = 0.0045$, $*P < 0.05$, $n = 4-6/\text{group}$. NS, not significant. Veh, vehicle; WT, wild type,



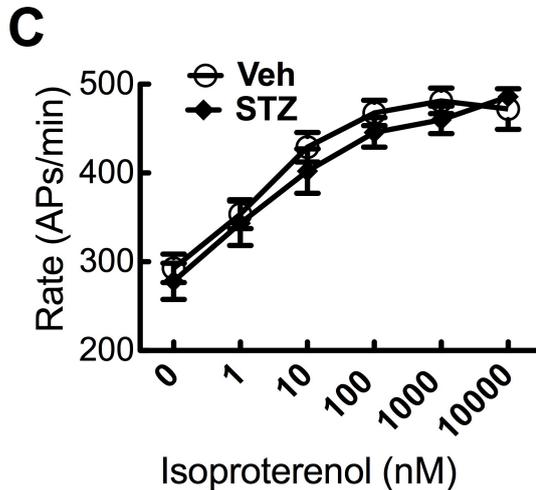
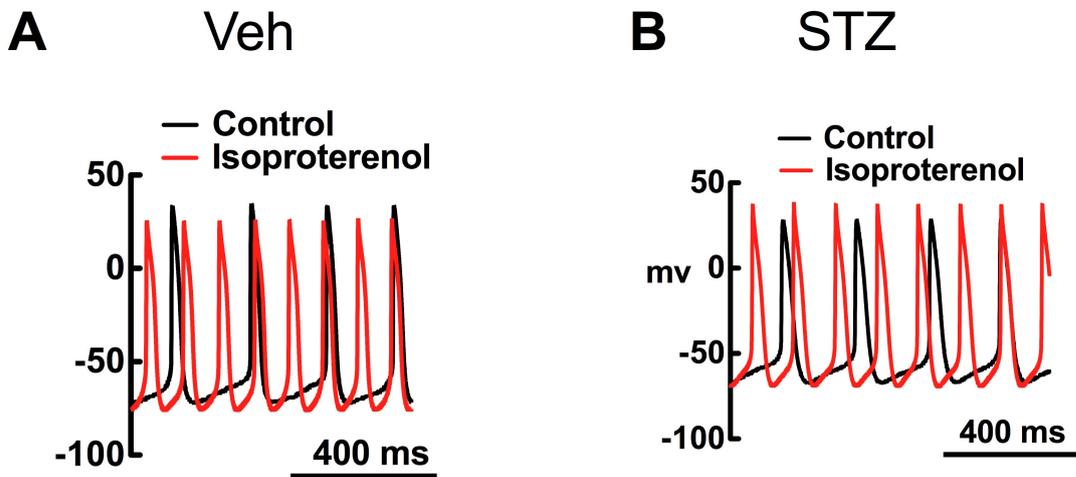
Supplemental Figure 7

Comparable left ventricular contractile function and chamber volumes in non-diabetic (2.1 ± 0.3 days after MI) and diabetic mice (2.0 ± 0.4 days after MI). (A) Ejection fraction (EF), (B) end diastolic volumes (EDV) and end systolic volumes (ESV) after MI in Veh- and STZ-treated WT mice, STZ-treated MM-VV, AC3-I, *Ncf1*^{-/-} or WT + Mito mice. Overall $P = 0.6$ for EF, overall $P = 0.6$ for EDV and 0.4 for ESV, $n = 4-11$ /group. Veh, vehicle; WT, wild type; Mito, MitoTEMPO



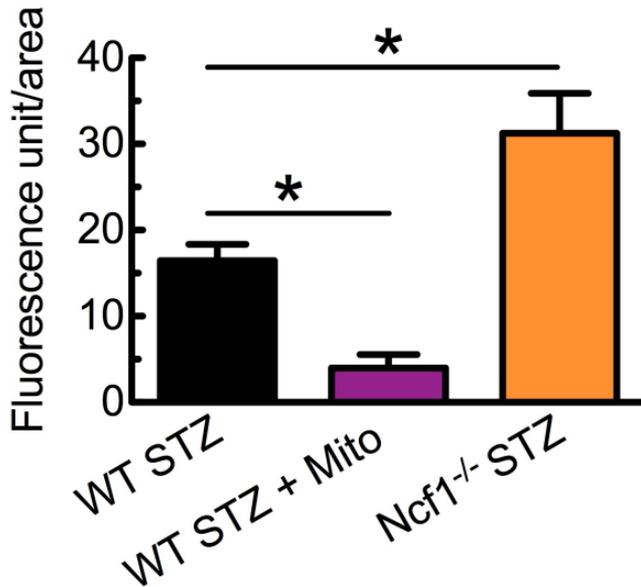
Supplementary Figure 8

Increased SAN apoptosis after MI in STZ-treated WT mice, but not MM-VV or AC3-I mice. Representative immunofluorescence images of TUNEL staining in SAN from WT mice treated with Veh + MI or STZ + MI, MM-VV and AC3-I with STZ + MI (Blue, DAPI; Green, HCN4; Red, TUNEL). Scale bars: 50 μ m. Overall $P < 0.0001$, $***P < 0.001$, $n = 4-8$ /group. Veh, Vehicle; WT, wild type



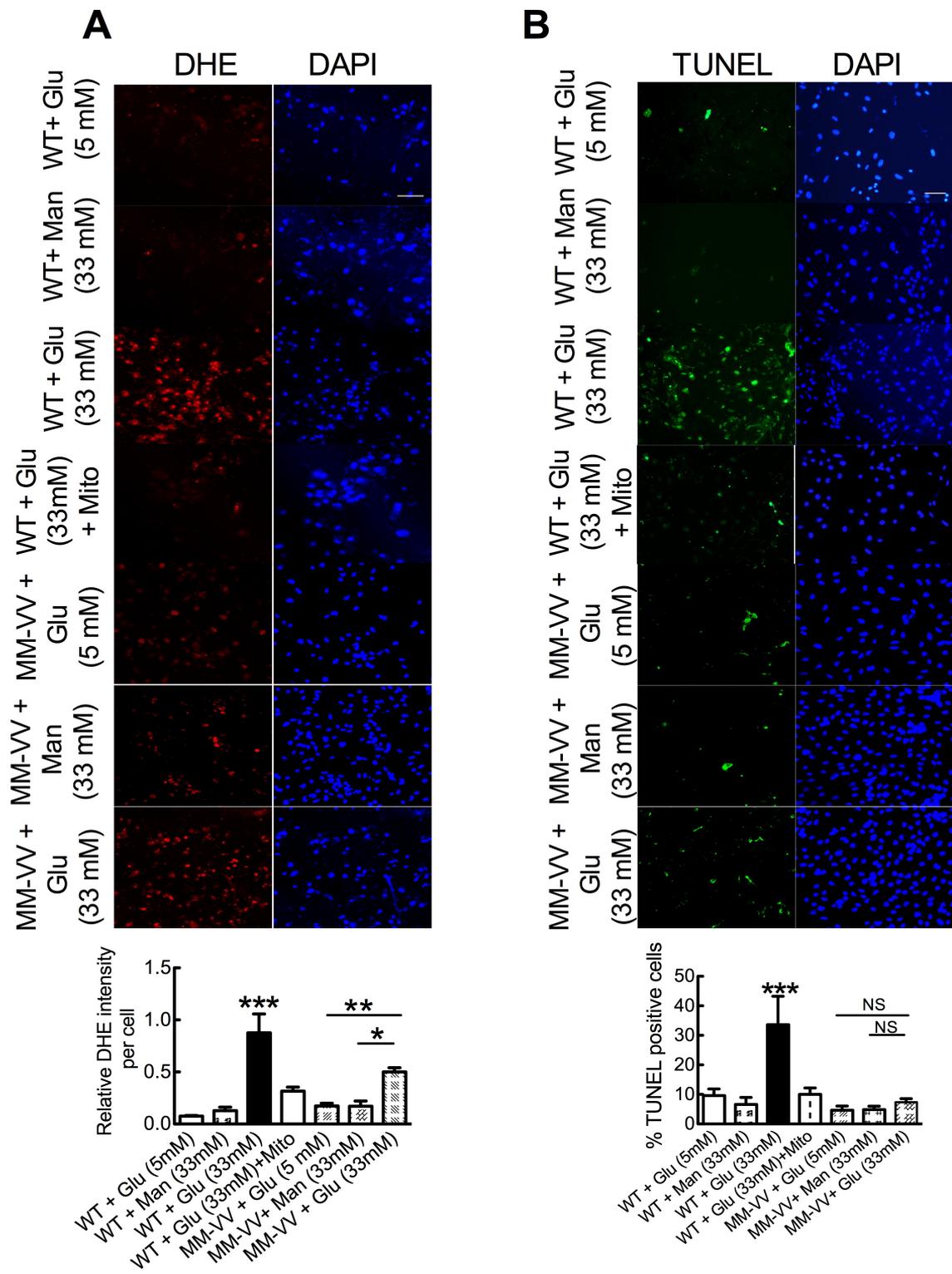
Supplemental Figure 9

STZ does not affect spontaneous or isoproterenol-stimulated automaticity in isolated SAN cells. (A, B) Representative tracings of spontaneous action potentials from SAN cells isolated from Veh- and STZ-treated mice with or without isoproterenol. (C) Summary data of action potential (AP) rates of SAN cells. $P > 0.05$ between Veh- and STZ-treated group at baseline and after isoproterenol at all doses, $n = 7-10$ cells/group. Veh, vehicle



Supplemental Figure 10

Decreased ROS in SAN from MitoTEMPOL-treated mice. DHE staining in SAN from WT mice treated with Mito + STZ, STZ alone and *Ncf1*^{-/-} mice treated with STZ. Overall $P = 0.0003$, $*P < 0.05$, $n = 3-5/\text{group}$. WT, wild type; Mito, MitoTEMPO



Supplemental Figure 11

Hyperglycemia triggers mitochondrial ROS generation and apoptosis in WT mice but not apoptosis in MM-VV mice. (A) DHE staining for ROS in cultured WT neonatal cardiomyocyte treated with glucose (Glu), Glu+Mito (MitoTEMPO, 1 mM) or mannitol (Man) and in MM-VV mice treated with glucose (Glu) or mannitol (Man). (B) TUNEL staining in the same experimental groups as in (A). For both panels, overall $P < 0.0001$, $***P < 0.001$ between WT + Glu (33mM) group and all other groups, $**P < 0.01$ and $*P < 0.05$ by Neuman-Keuls test as post hoc study. NS, not significant, $n = 3-5$ assays/group. WT, wild type

	Vehicle	STZ
Body weight (g)	25 ± 0.5	21 ± 0.8*
Heart weight/tibia length (mg/mm)	8.5 ± 0.6	9.2 ± 0.5
Lung weight/tibia length (mg/mm)	8.1 ± 0.4	8.8 ± 0.6
BUN (mg/dl)	24 ± 4	26 ± 1
CO₂ (mEq/l)	15 ± 3.5	14 ± 1.5

Supplemental Table 1. Body weights, organ weights and blood chemistry of vehicle and STZ-treated mice after MI. Body weights (* $P < 0.001$, $n = 24-25$ /group), lung weights normalized to tibia length ($P = 0.4$, $n = 6-7$ /group), and heart weights normalized to tibia length ($P = 0.4$, $n = 6-7$ /group) compared between vehicle- and STZ-treated mice. Serum BUN ($P = 0.5$, $n = 6-8$ /group) and bicarbonate levels ($P = 0.6$, $n = 5-9$ /group) compared between vehicle- and STZ-treated mice.

Echocardiography Parameters	Veh + MI	STZ + MI	P
Ejection Fraction (%)	38.8 ± 0.07	34.0 ± 0.09	0.67
End Diastolic Volumes (ml)	60.8 ± 3.6	63.1 ± 9.5	0.82
End Systolic Volumes (ml)	36.1 ± 5.1	45.5 ± 10.5	0.41
Stroke volume (ml)	24.7 ± 3.2	18.0 ± 2.2	0.11
Heart Rate (BPM)	570 ± 33.7	398 ± 51	0.01*
Cardiac output (ml/min)	14.3 ± 2.1	7.1 ± 1.5	0.019 *
Left ventricular mass (mg)	86.4 ± 6.6	80.2 ± 7.0	0.53

Supplemental Table 2. STZ-treated mice had decreased heart rate and cardiac output, but similar contractile function, left ventricular volumes and mass compared to vehicle-treated mice after MI. P values for comparisons between Veh- and STZ-treated mice after MI, n = 7-9/group. Veh, vehicle.

Patient Characteristics	MI + DM (n=5)	MI (n=5)	P
Age (year)	64 ± 7	71 ± 2	0.03
Males (%)	75	60	
Ejection fraction (%)	41 ± 6	35 ± 6	0.5
Type II DM (%)	100	n/a	
Insulin (%)	75	n/a	
Glucose (mg/dl)	260 ± 56	121 ± 13	0.04
Hemoglobin A1C (%)	8.0 ± 0.5	n/a	
Pacemaker (%)	25	0	
ACE inhibitor (%)	80	100	
β-Blocker (%)	100	100	

Supplemental Table 3: Patient characteristics for right atrial tissues shown in Figure 3. ACE,

Angiotensin Converting Enzyme.

Cell Type	Source	$R_{gap, longitudinal}$	$R_{gap, transverse}$
Atrial	Courtemanche et al. 1998 [*]	1.5 Ωcm^2	15.0 Ωcm^2
Peripheral SAN	Kurata et al. 2008 [#]	<i>Equation 2</i>	$R_{gap, long}$
Central SAN	Kurata et al. 2002 [§]	<i>Equation 2</i>	$R_{gap, long}$
Block zone	Butters et al. 2010 [‡]	4000 Ωcm^2	$R_{gap, long}$
Inexcitable cell	Morita et al. 2009 [†]	4000 Ωcm^2	$R_{gap, long}$

Supplemental Table 4. Mathematical models for different regions of intact SAN

^{*}Courtemanche et al. *Am J Physiol.* 1998;275:H301-321.

[#]Kurata et al. *Biophys J.* 2008;95:951-977 with sodium channel conductance = 1.8523×10^{-6} nS/pF.

[§]Kurata et al. *Am J Physiol Heart Circ Physiol.* 2002;283:H2074-2101.

[‡]Butters et al. *Circ Res.* 2010;107:126-37. Block zone modeled as coupled region of inexcitable cells with passive leak conductance ($E_{leak} = -70$ mV).

[†]Morita et al. *Am J Physiol Heart Circ Physiol.* 2009;297:H1594.

State variable	Definition	Initial value
m	Na ⁺ current activation gate	0.004147463955
h	Na ⁺ current inactivation gate	0.9421032891
j	Na ⁺ current slow inactivation gate	0.9319148046
d	L-type Ca ²⁺ current activation gate	0.0001793553646
f	L-type Ca ²⁺ voltage-dependent inactivation gate	0.8466729054
f_{ca}	L-type Ca ²⁺ calcium-dependent inactivation gate	0.5727907784
X_r	Rapidly activating K ⁺ current activation gate	0.02303434883
X_s	Slowly activating K ⁺ current activation gate	0.03616188931
oa	Transient outward K ⁺ current activation gate	0.03436021906
oi	Transient outward K ⁺ current inactivation gate	0.9987671846
ua	Ultrarapid rectifier K ⁺ current activation gate	0.006237057018
ui	Ultrarapid rectifier K ⁺ current inactivation gate	0.9891443725
u	Ryanodine receptor Ca ²⁺ release activation gate	4.537711703e-16
v	RyR Ca ²⁺ release inactivation gate	1.0
w	RyR Ca ²⁺ release inactivation gate	0.9990895327
$[Ca^{2+}]_i$	Ca ²⁺ concentration in myoplasm (mM)	0.0002599348619
$[Ca^{2+}]_{JSR}$	Ca ²⁺ concentration in junctional SR (mM)	0.4298392427
$[Ca^{2+}]_{NSR}$	Ca ²⁺ concentration in network SR (mM)	1.960512954
$[Na^+]_i$	Na ⁺ concentration in myoplasm (mM)	15.19468146
$[K^+]_i$	K ⁺ concentration in myoplasm (mM)	134.9547735
V_m	Transmembrane potential (mV)	-79.01495665

Supplemental Table 5. Initial conditions for state variables in mathematical model of atrial action potential*. *Single cell was paced to steady-state at cycle length = 300 ms. Model equations for human atrial cell model are found in original publication (1). SR, sarcoplasmic reticulum.

State	Definition	Initial value	Initial value
variable		(Central)	(Peripheral)
<i>m</i>	Na ⁺ current activation gate	-	0.0690372072
<i>h</i>	Na ⁺ current inactivation gate	-	0.2223618089
<i>j</i>	Na ⁺ current slow inactivation gate	-	0.01430146964
<i>d</i>	L-type Ca ²⁺ current activation gate	0.0006779558663	4.66775128e-05
<i>f</i>	L-type Ca ²⁺ voltage-dependent inactivation gate	0.6621880012	0.459162836
<i>f_{ca}</i>	L-type Ca ²⁺ calcium-dependent inactivation gate	0.5710787057	0.4667391842
<i>X_{r,f}</i>	Rapidly activating K ⁺ current fast activation gate	0.3160790736	0.5254608664
<i>X_{r,s}</i>	Rapidly activating K ⁺ current slow activation gate	0.6156126509	0.6219289422
<i>X_{r,i}</i>	Rapidly activating K ⁺ current inactivation gate	0.847082724	0.933848794
<i>n</i>	Slowly activating K ⁺ current activation gate	0.0552203568	0.07350047887
<i>q</i>	Transient outward K ⁺ current activation gate	0.5239671546	0.5779873691
<i>r</i>	Transient outward K ⁺ current inactivation gate	0.005795549972	0.00199964719
<i>dt</i>	T-type Ca ²⁺ current activation gate	0.005151304518	0.0003472296223
<i>ft</i>	T-type Ca ²⁺ current inactivation gate	0.2498661863	0.3016670853
<i>qa</i>	Sustained inward current activation gate	0.4591365202	-
<i>qi</i>	Sustained inward current inactivation gate	0.3860293202	-
<i>y</i>	Pacemaker (funny) current activation gate	0.02836884815	0.01632643918

$[Ca^{2+}]_i$	Ca^{2+} concentration in myoplasm (mM)	0.00023567935	0.0002461860556
$[Ca^{2+}]_{JSR}$	Ca^{2+} concentration in junctional SR (mM)	0.3477078758	0.219459533
$[Ca^{2+}]_{NSR}$	Ca^{2+} concentration in network SR (mM)	2.518529555	4.245147564
$[Na^+]_i$	Na^+ concentration in myoplasm (mM)	9.211637929	7.679366896
$[K^+]_i$	K^+ concentration in myoplasm (mM)	140.1632028	141.5993186
V_m	Transmembrane potential (mV)	-57.88001647	-74.11420057

Supplemental Table 6. Initial conditions for state variables in mathematical models of central and peripheral SAN cells*. **Single cell underwent spontaneous activity for 10 seconds. Model equations for rabbit central and peripheral node cells are found in original publication (2). SR, sarcoplasmic reticulum*

Supplemental equations for mathematical modeling as below:

Capacitive membrane area (cm^2)

x is distance in cm .

$$A_{cap} = 110 \times 10^{-6} - 45 \times 10^{-6} \left(\frac{1}{1+\exp(-4(x-7.0))} + \frac{1}{1+\exp(4(x-9.0))} \right) \quad [1]$$

Gap junction resistance (Ωcm^2)

$$R_g = 110 \left(\frac{1}{1+\exp(-4(x-7.0))} + \frac{1}{1+\exp(4(x-9.0))} \right) - 108.5 \quad [2]$$

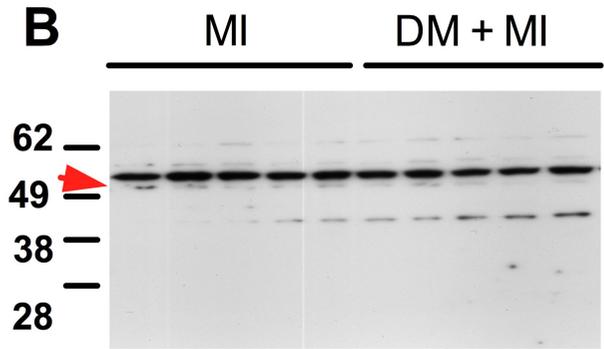
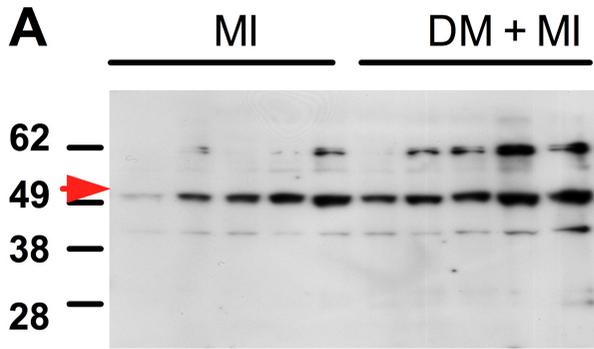
Conductance of fast sodium current (mS/cm^2)

$$\bar{g}_{Na} = 3.7 \times 10^{-6} - 1.85 \times 10^{-6} \left(\frac{1}{1+\exp(-5(x-6.5))} + \frac{1}{1+\exp(5(x-9.5))} \right) \quad [3]$$

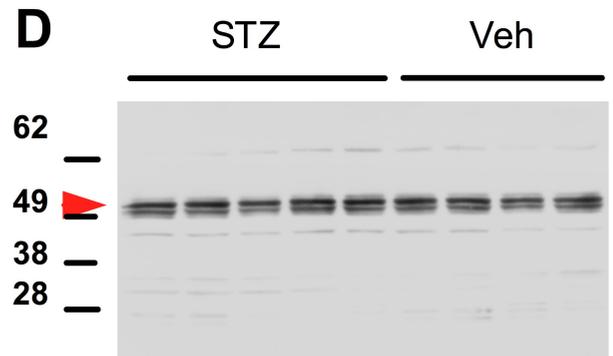
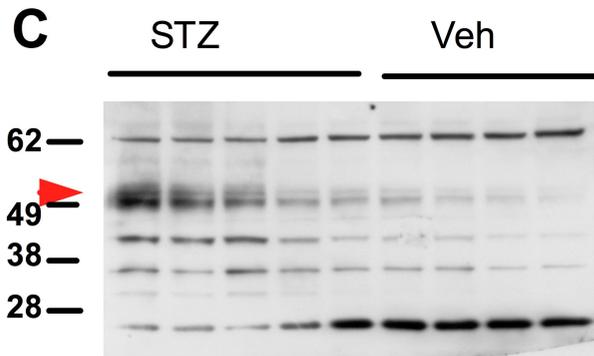
References:

1. Courtemanche, M., Ramirez, R.J., and Nattel, S. 1998. Ionic mechanisms underlying human atrial action potential properties: insights from a mathematical model. *American Journal of Physiology* 275:H301-321.
2. Kurata, Y., Matsuda, H., Hisatome, I., and Shibamoto, T. 2008. Regional difference in dynamical property of sinoatrial node pacemaking: role of na^+ channel current. *Biophys J* 95:951-977.

Full unedited gel for Figure 3A

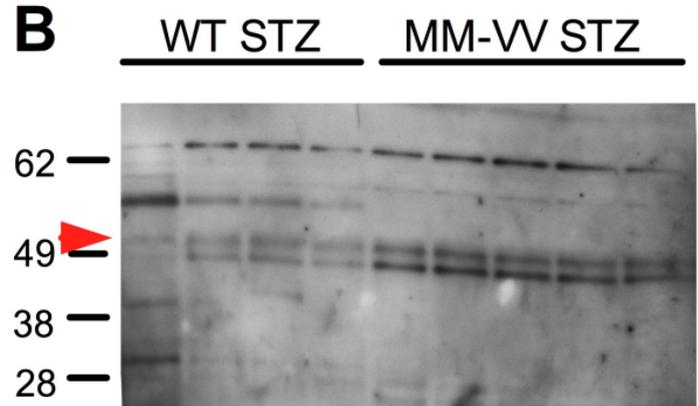
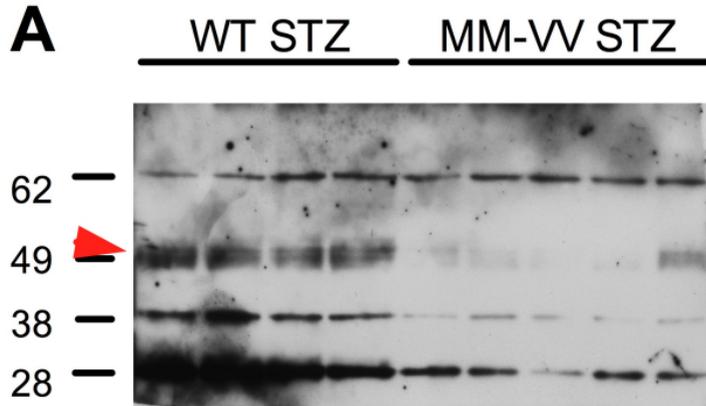


Full unedited gel for Figure 3B



Western blot for Figure 3 A, B. ox-CaMKII (A, C) and CaMKII (B, D) staining of human and mice right atrial samples. Red arrow denotes ox-CaMKII or CaMKII.

Full unedited gel for Figure 4A



Western blot for Figure 4A. ox-CaMKII (A) and CaMKII (B) staining of right atrial samples from WT and MM-VV mice. Red arrow denotes ox-CaMKII or CaMKII.